# **BME Chemical Toolkit 1.0**

Created by Patrick Chirdon 2020 with the support of Dr.Douglas Goetz, Dr.Sumit Sharma, Dr.Colin Kruse, and Athena Ghiasi at Ohio University

Requires Python

Spark

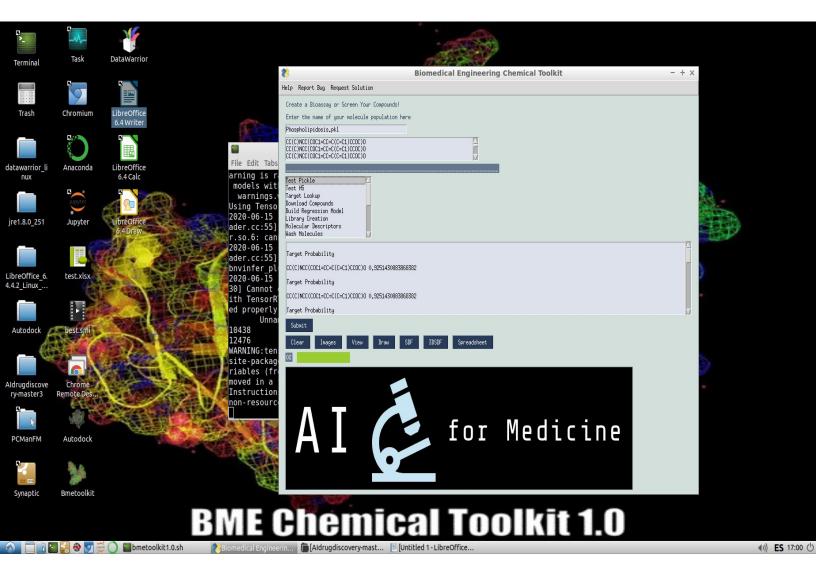
Java

Anaconda

Virtualbox

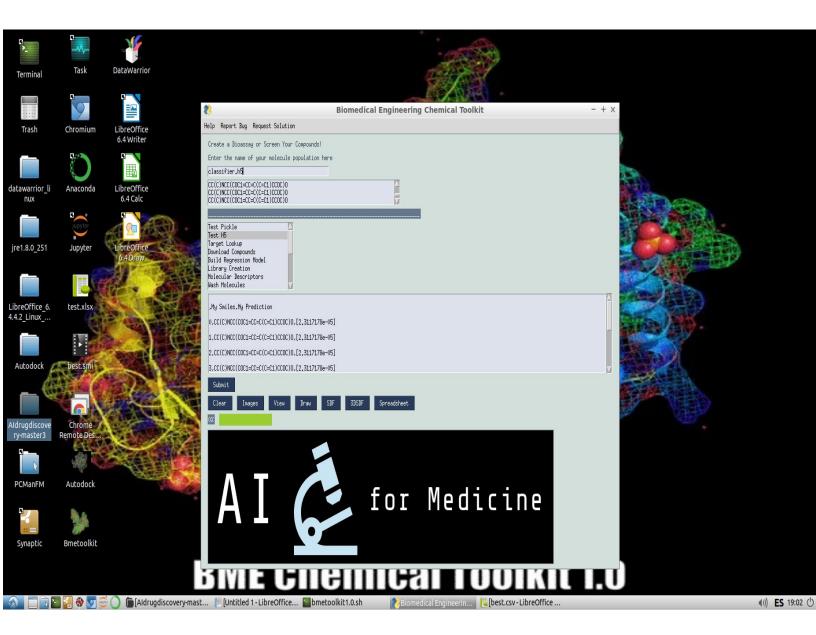
## 1.Test Pickle

Input the pickle name into the first box. The file must end in .pkl. Then input a list of smiles into the second box, click test pickle and then click submit. The output is a target probability.



#### **2. Test H5**

Type the name of the .h5 in the first box. Type a list of SMILES into the second box and click submit. The output is a probability.



# 3. Target Lookup

Type in the name of the protein target into the first box.

The output will be a CHEMBL ID. ie

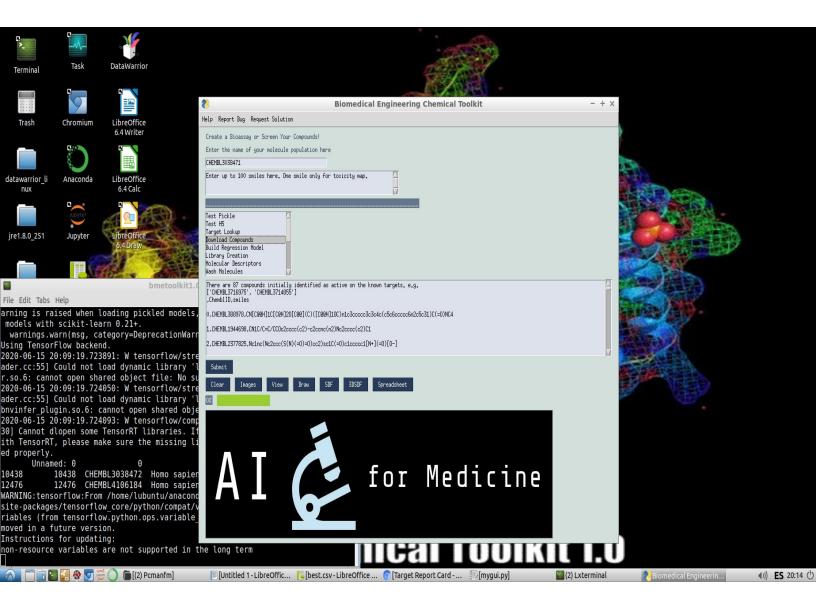
 $C[C@@H](C(=O)N[C@@H](CO)C(=O)N1CCC[C@H]1C(=O)N[C@@H] \\ (CCCN=C(N)N)C(=O)N[C@@H](CCCCN)C(=O)O)NC(=O)[C@H](CC2=CN=CN2)NC(=O)[C@H] \\ (CC3=CN=CN3)N$ 

Unnamed: 0		0	1	2	
10435	10435	CHEMBL	3038469	Homo sapiens	CDK2/Cyclin A
10436	10436	CHEMBL	3038470	Homo sapiens	CDK2/Cyclin A1
10483	10483	CHEMBL	3038517	Homo sapiens	CDK2/CDK4
12444	12444	CHEMBL4	4106152	Homo sapiens	CDK2/Cyclin O
12445	12445	CHEMBL4	4106153	Homo sapiens	CDK2/Cyclin A
12477	12477	CHEMBL4	4106185	Mus musculus	CDK2/CDK4
12478	12478	CHEMBL <sup>2</sup>	4106186	Mus musculus	CDK2/CDK9



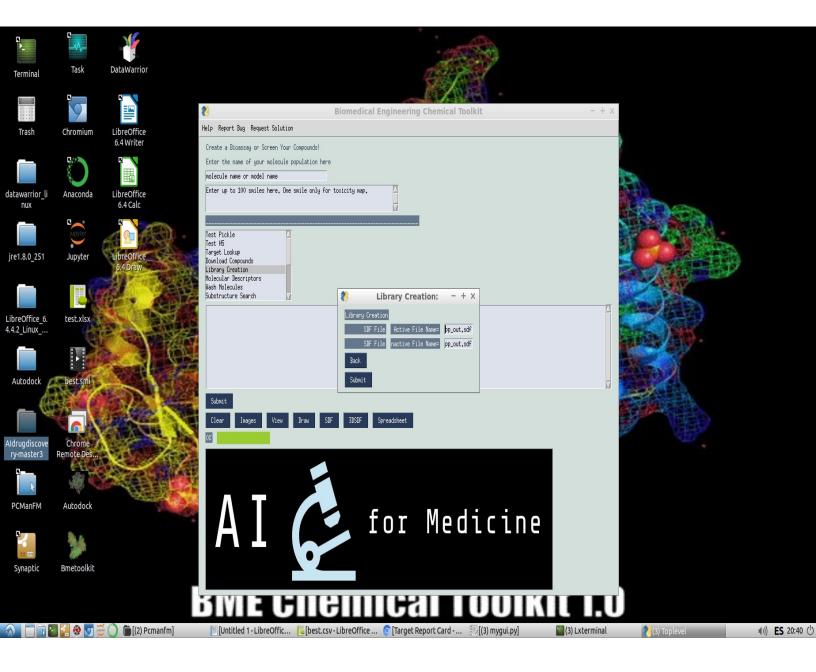
## 4. Download Compounds

Input the CHEMBL ID into the first box and click submit. The output will be a list of smiles and their corresponding CHEMBLID for the compound



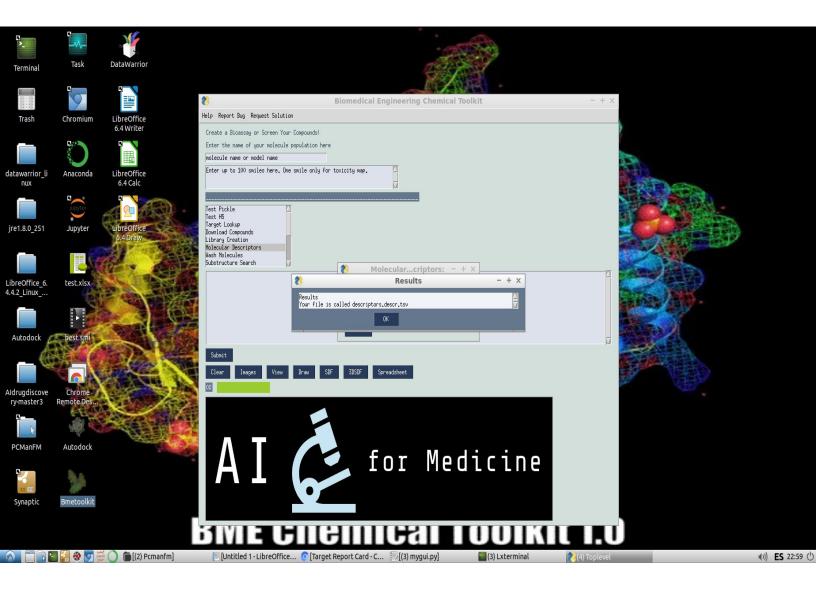
## 5. Library Creation

Create a library of compounds labeled active, 1, or inactive, 0 as a SDF file. Input an active file name containing compounds that are known to bind a target and input an inactive file name containing compounds that do not bind (could also be just random compounds)



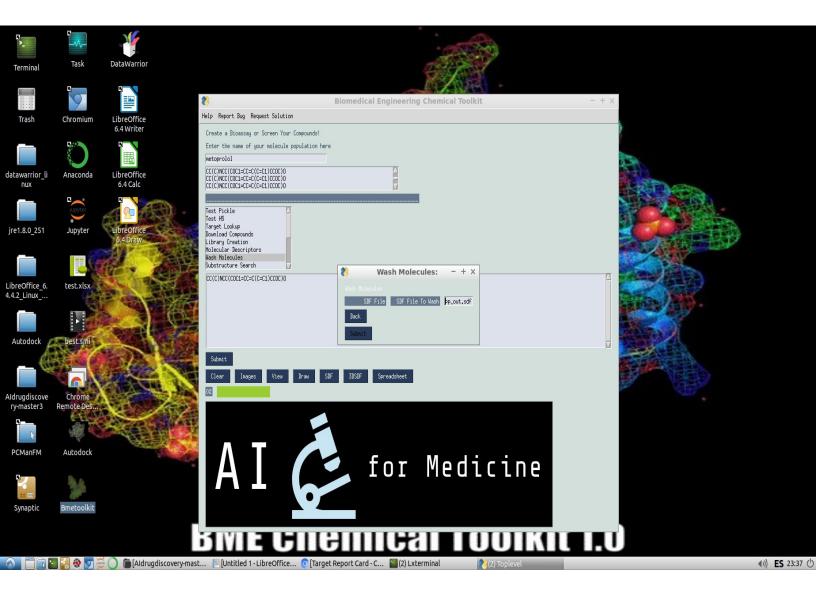
## 6. Molecular Descriptors

Input a SDF file. The output file will be called descriptors.descr.tsv. The output will be an excel file with chemical properties.



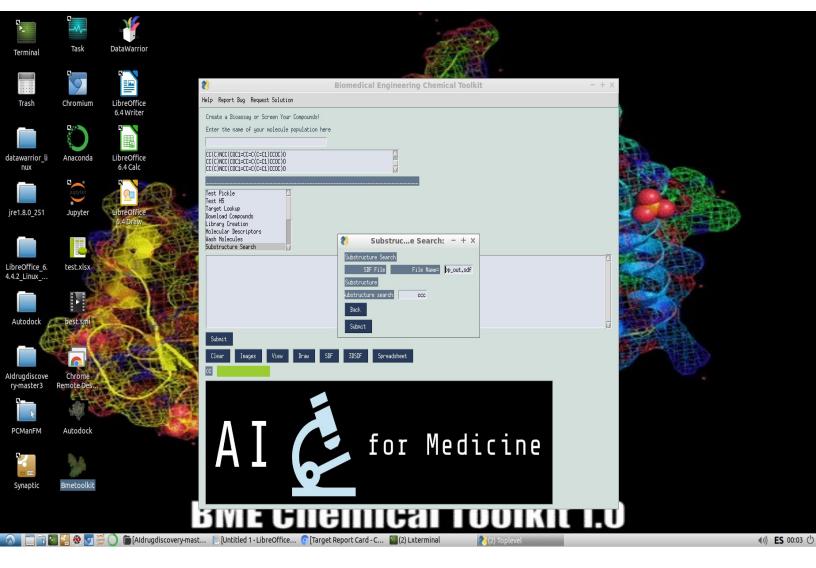
### 7. Wash Molecules

Input an SDF molecule. The program will remove some metal atoms and ions, remove small fragments, adjust formal charges, and rebalance hydrogen counts.



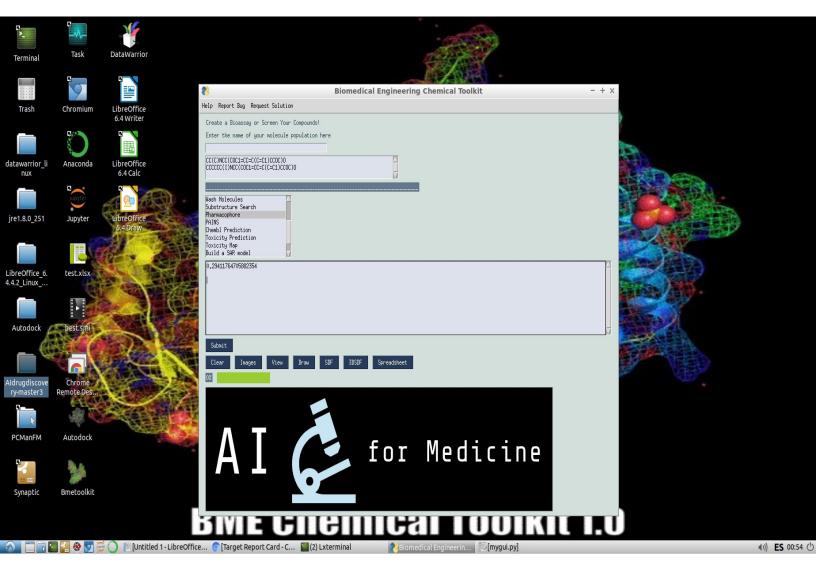
#### 8. Substructure Search

For a substructure search input a SDF file and search for a substructure. The results output will be a list of molecules containing the substructures.



## 9. Pharmacophore

Input two smiles and click pharmacophore. The output will be the pharmacophore similarity. 1 means that the molecules are the same. The scale is from 0 to 1.



#### 10. PAINS

#### PAN Assay Interference

The program will tell you what functional groups for each compound were responsible for a positive PAINFUL test result. The program also tells you the fraction of SP3 hybridized carbons. Compounds with scores > .47 are more selective binders. Note that double bonds reduce the fraction of sp3 hybridization, as they make the compound more flat. See Escape from flatland: increasing saturation as an approach to improving clinical success. Pains are defined as follows:

Doveston R, et al. A Unified Lead-oriented Synthesis of over Fifty Molecular Scaffolds. Org Biomol Chem 13 (2014) 859D65. doi:10.1039/C4OB02287D

Jadhav A, et al, Quantitative Analyses of Aggregation, Autofluorescence, and Reactivity Artifacts in a Screen for Inhibitors of Thiol Protease. J Med Chem 53 (2009) 37D51. doi:10.1021/jm901070c

#### 11.Chembl Prediction

**Chembl Target Prediction** 

Multiclass Classifier

Number of unique targets 560

Ion channel 5

kinase 96

nuclear receptor 21

**GPCR 180** 

Others 258

accuracy .87

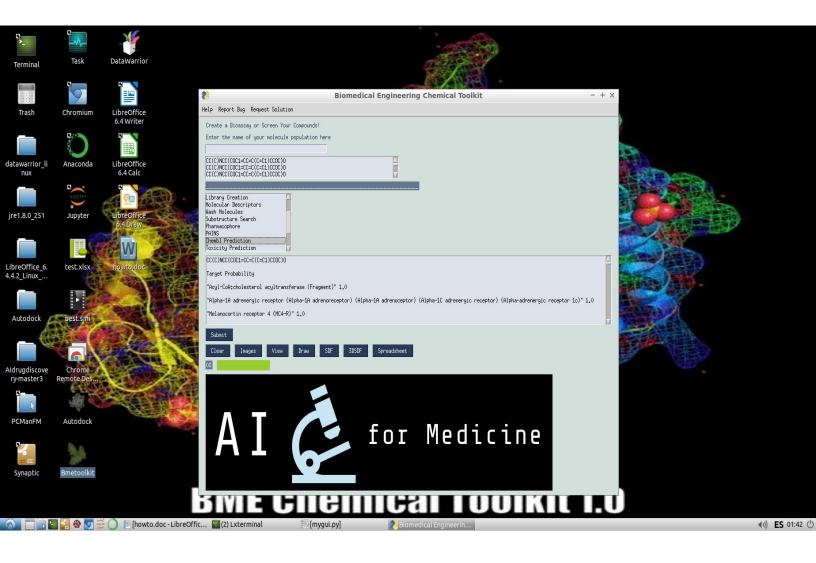
auc .92

sensitivity .76

specificity .92

#### precision .82

225/225 100% correct mechanism on independent test set. Note-- 1 is considered positive and zero is negative for a given target.



## 12. Target Prediction

Support Vector Machine

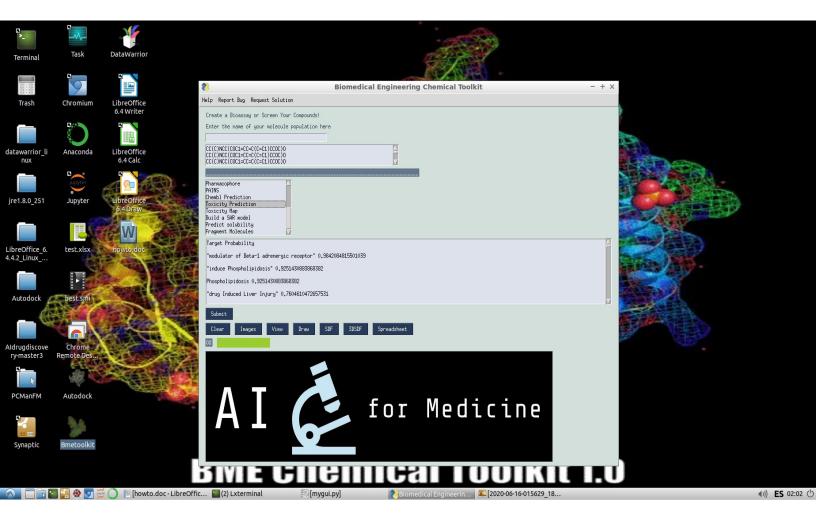
46 models. Metrics for test data:

accuracy 97.59 +/- 2.41

sensitivity 91.9 +/- 8.1

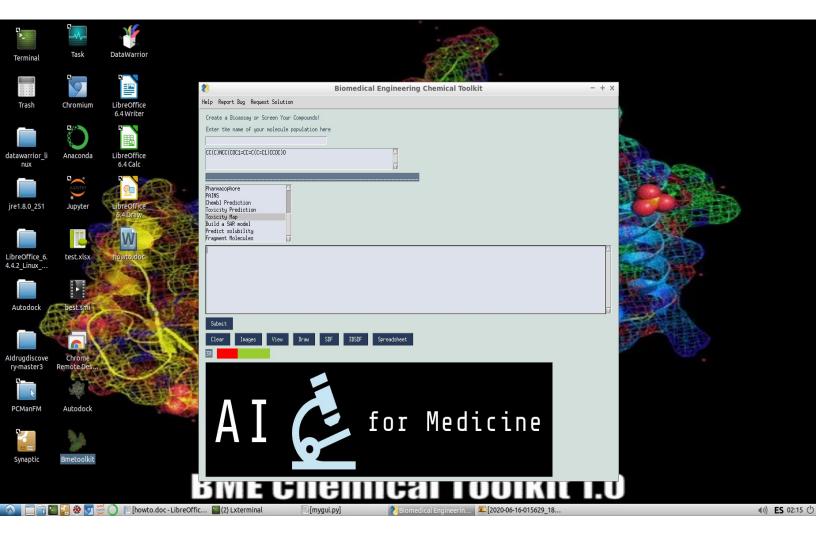
specificity 98.6 +/- 1.4

215/247 87% correct mechanism on independent test set.



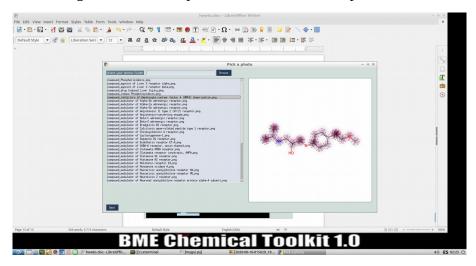
## 13. Toxicity Map

Input One SMILE. The output will be 46 toxicity maps for the 46 models available from toxicity prediction.

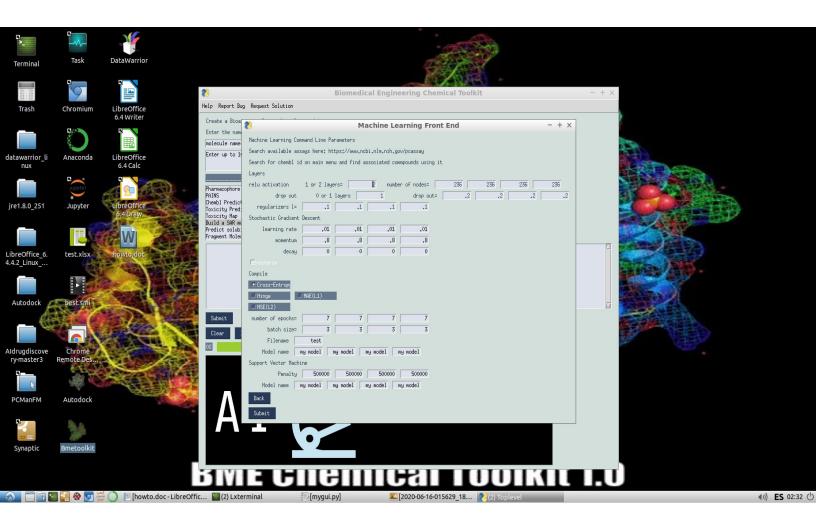


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Click Images to see the output. Pink marks inactive parts of the molecule, green marks active parts



#### 14. Build a SAR Model



Toggle the parameters. There are 4 models for the SVM and 4 models for the neural network
cross entropy- default loss function for binary classification problems. Summarizes the average difference between the actual and predicted probability.
hinge- alternative to cross entropy binary classification developed with SVM models used with support vector machine models
mse-default loss to use for regression problems. calculated as the average of the squared differences between the predicted and actual values
mae-for regression problems. used in cases where there are outliers, average of the absolute difference between actual and predicted values

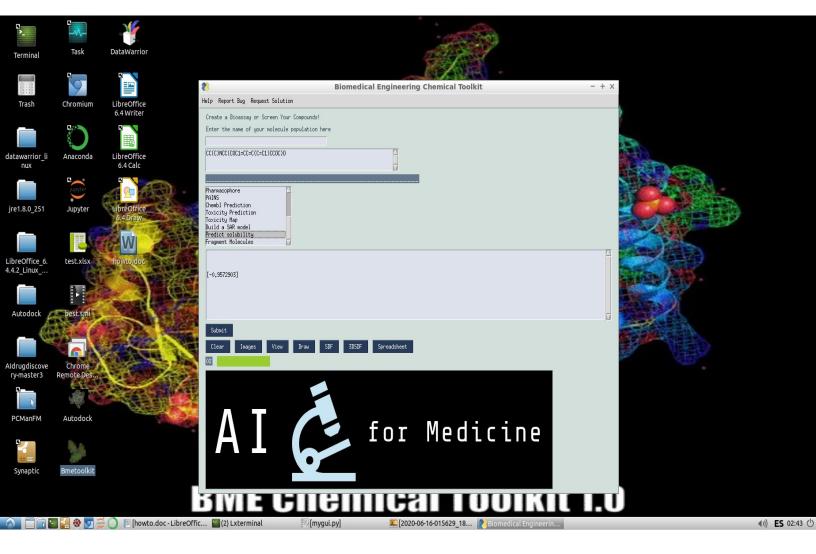
# 15. Solubility

Solubility

Predicts log S. Log S greater than -4 is soluble.

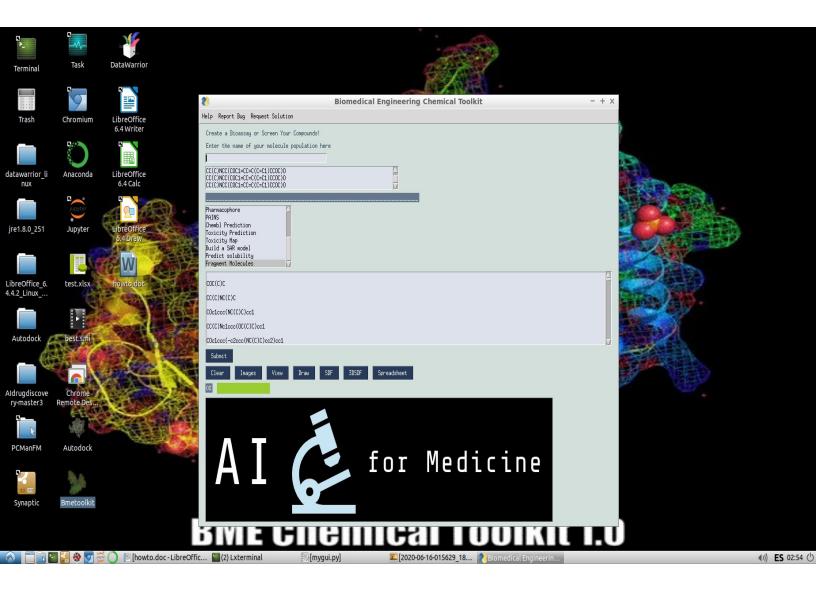
Root mean square error of 1.27 on a scale from -4 to 4.

linear regression



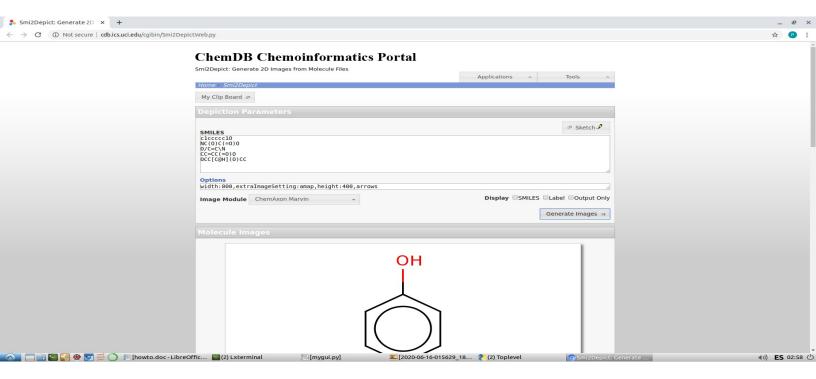
### 16. Fragmenter

Input a list of smiles. These will be recombined into new combinations. When you take the lowest energy ligands from a docking program and recombine these there may be some compounds that bind with lower energy than the original.

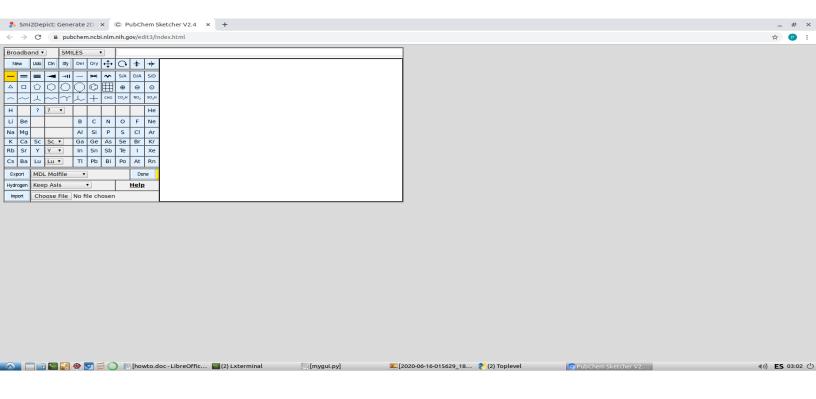


#### **17. View**

#### Input a list of SMILES and click Generate to view

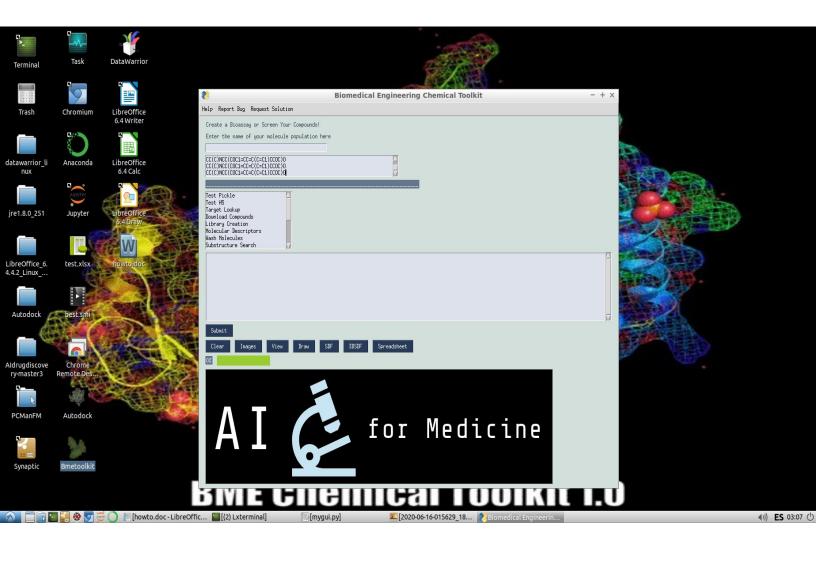


#### 18. Draw



## 19. SDF

Input a list of molecules. Click SDF. The output will be a 2D SDF file called output.sdf

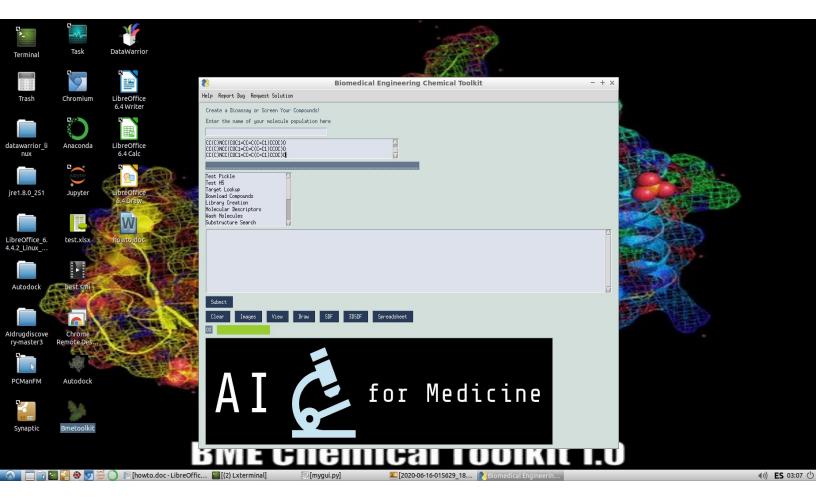


### 20.3D SDF

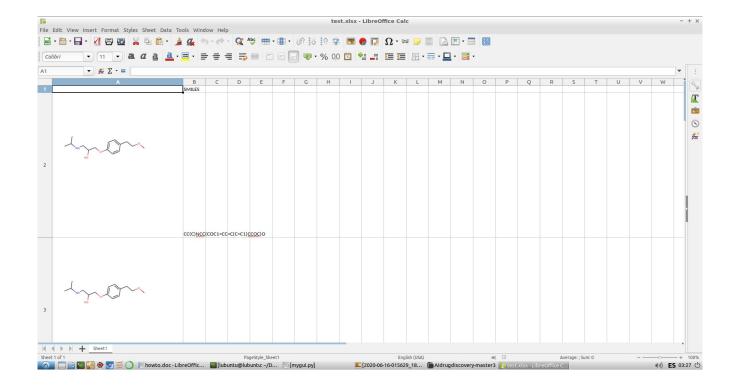
Input a list of SMILES and the output will be a 3D SDF file called output.sdf



# 21. Spreadsheet



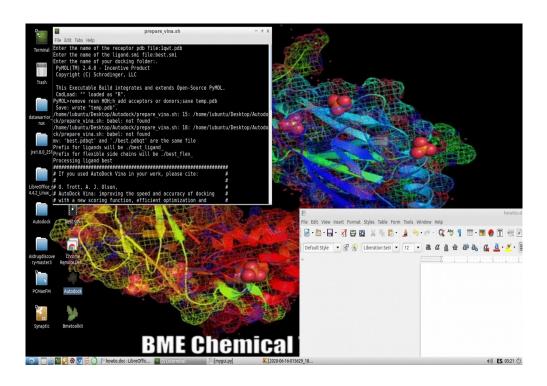
Input a list of SMILES. next to them.	The output will be a spread	lsheet called test.xlsx wi	th pictures and SMILES



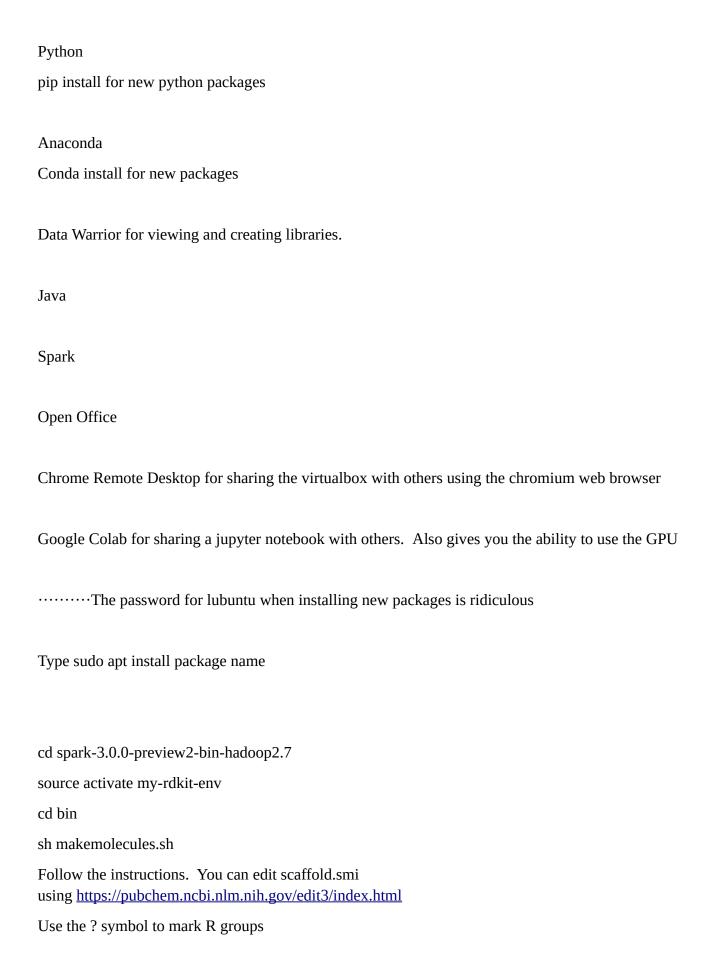
# 22. Prepare Autodock VINA

- 1. Enter the name of the receptor pdb
- 2. Enter the name of the ligand.smi file
- 3. Enter the docking folder as

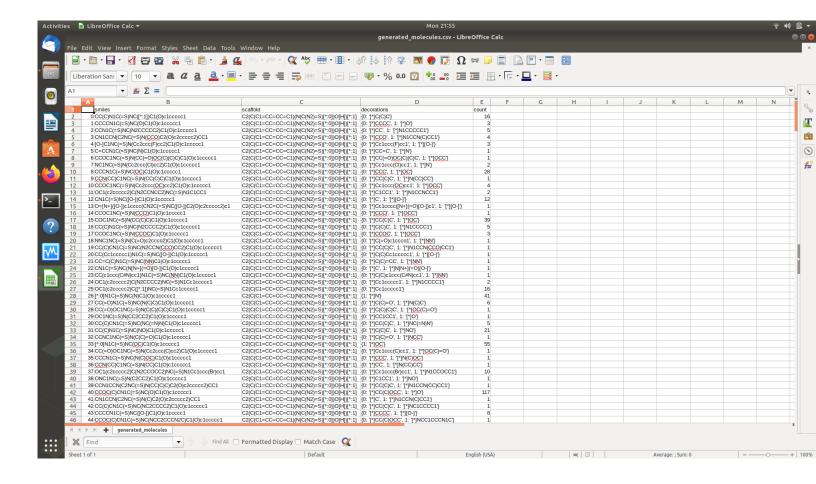
The output will be a separate directory for each receptor ligand complex with a log file.



There will be an output file called results table.csv in the autodock folder that will contain the docking energy in kJ for each of the compounds. The lower the energy the better the binder.

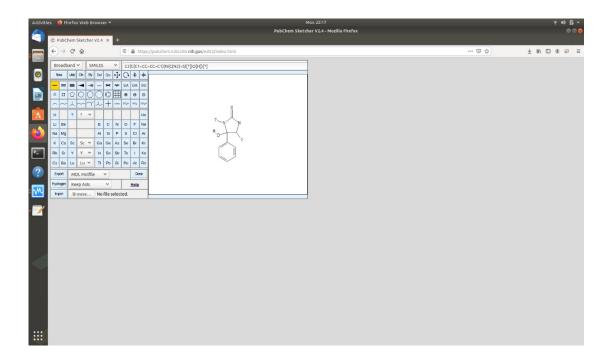


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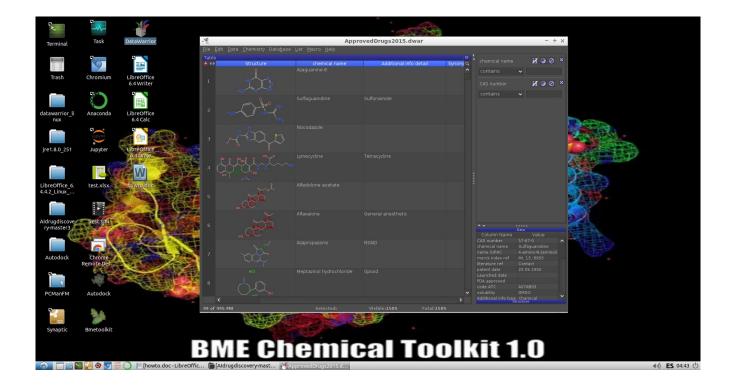
The output will be a csv file called generated molecules.csv that contains the generated SMILES and the scaffold it was generated from

Here is an example scaffold.

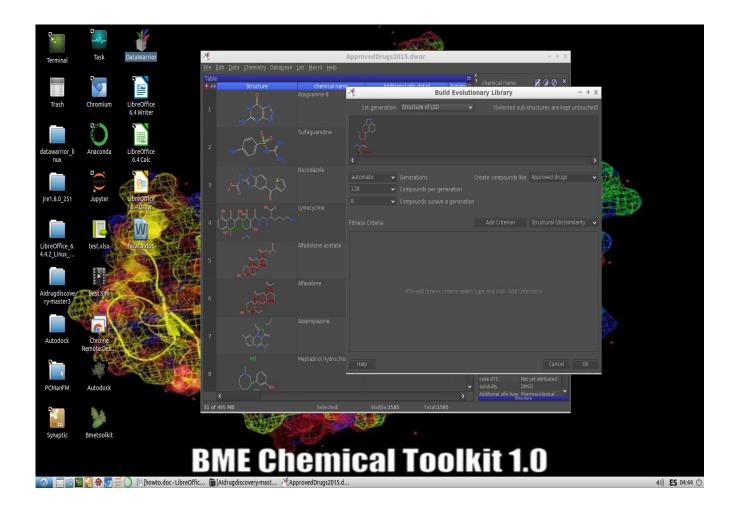


Other useful packages pre installed fpocket, openbabel, and pymol. See documentation online

Data Warrior Library View allows you to store and edit libraries

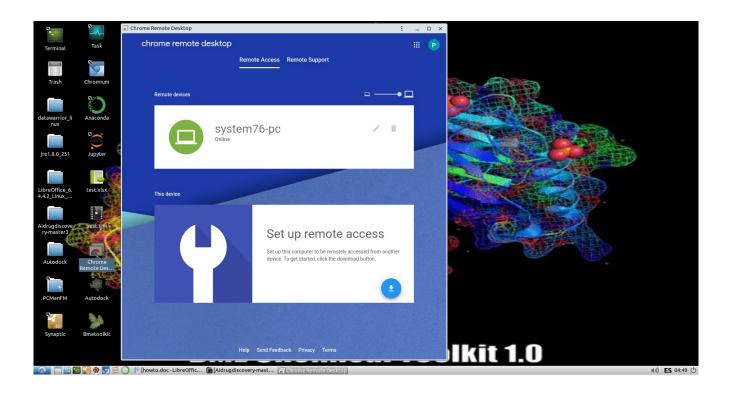


Generate an Evolutionary Library



Calculate a Drug Score under Macro, Calculate Properties

Chrome Remote Desktop allows you to share your screen remotely with someone through the browser



open the shell type babel and you can convert between formats

fpocket

Allows you to find binding sites to use as docking coordinates for autodock

PyMOL

Allows you to view images

